

# Chitosan in bio-based food contact materials

**This is a paper for discussion.**

**This does not represent the views of the Committee and should not be cited.**

## Introduction

1. In May 2020, a scoping paper entitled “Alternatives to conventional plastics for food & drinks packaging” (TOX/2020/24), which introduced some of the possible toxicological hazards associated with the use of bio-based food contact materials (BBFCMs), was presented to the Committee on Toxicity of Chemicals in Food, Consumer Products and the Environment (COT). Subsequently, a proposed list of BBFCMs for health risk assessment was presented to the COT in February 2021 (TOX/2021/01); this included BBFCMs containing chitosan.
2. Attached at Annex A to this paper is a draft position paper on chitosan in bio-based food contact materials. This is a second draft which has been revised following the Committee’s discussion at the September 2022 meeting, when the first draft of the position paper ([TOX/2022/45](#)) was presented.
3. At the COT meeting held in July 2023, the Committee considered a paper entitled “draft assessment of the Codex report on food allergen thresholds” (TOX/2023/35). At this meeting, the Committee agreed with how the assessment has been undertaken by the COT subgroup and with the contents and key conclusions reached by the COT subgroup. However, Members requested that a sense check on clinical terminology is carried out on Annex A of TOX/2023/35, and that paragraph 17 of Annex A of TOX/2023/35 should be rephrased to clarify the conclusions.
4. The changes made to the draft position paper (highlighted in yellow) therefore include additional context and background on 1) allergen reference doses (such as how the ED01 and ED05 values are derived, and recent

recommendations made by the Joint FAO/WHO Expert Consultation on risk assessment of food allergens; paragraphs 28-30); 2) the other allergenic proteins which are present in crustacea in addition to tropomyosin (paragraph 10); and 3) how the migration limit of 10 mg/dm<sup>2</sup> in the Plastics Regulation was derived and clarification on whether it represents a maximum allowable limit (paragraph 42). Additional changes include the following:

- Clarification on whether the references to ‘shells’, ‘shellfish’, ‘shellfish allergy’, or ‘seafood allergy’ are referring to crustacea and/or molluscs (paragraphs 6-7, 9);
- Clarification that chitosan coatings are possible on food surfaces, and also the packaging itself (paragraph 13);
- The molecular weight of 10 kDa for chitooligosaccharides (COS) appears to be high, since other study authors note an average molecular weight of approximately 3.9 kDa or less (paragraph 26);
- Clarification that in the study of Waibel et al. (2011), only 10 of the 19 participants completed the study because only 10 individuals had met the inclusion criteria (paragraph 46); and,
- Regarding the evaluation of insect chitin, it should be clarified that this evaluation is on the whole mealworm preparation (not just the insect chitin), and therefore this specific evaluation is a more holistic assessment of allergy (paragraph 51).

5. At the COT meeting in September 2022, additional context was also requested by Members on how to interpret on life-cycle assessment (LCA) studies regarding chitosan in the environment. This was requested in order to help the Committee make any conclusions on the LCA studies which were described in the first draft of the position paper ([TOX/2022/45](#)). The information obtained has been presented below.

### **Life-cycle assessments (LCAs)**

6. SimaPro<sup>TM</sup> (developed by PRé Sustainability) is a widely used life cycle assessment (LCA) tool that enables users to evaluate the environmental impacts of products, processes, and services throughout their entire life cycle. While SimaPro can provide information on the potential release of particulates and other pollutants during a product's life cycle, the software does not directly assess degradation or particulate formation.

7. In SimaPro, users typically model a product or process as a series of interconnected stages, such as raw material extraction, transportation,

manufacturing, use, and end-of-life disposal. For each stage, the software provides data on a range of environmental impacts, including greenhouse gas emissions, energy use, water consumption, and waste generation. In terms of particulate emissions, SimaPro includes a database of environmental impact data that includes information on the release of particulate matter (PM) associated with specific materials, processes, and technologies. Users can also input data on the emission factors of different pollutants, including PM, to refine the accuracy of their LCA models. However, SimaPro does not specifically model the formation of particulates or other pollutants that may occur during a product's use or disposal phases. Users may need to supplement their LCA models with additional data or models that provide more detailed information on these processes.

8. There are generally agreed-upon inputs and outputs for conducting an LCA. The International Organization for Standardization (ISO) has developed a set of guidelines known as [ISO 14040:2006](#) which outlines the principles, framework, and requirements for conducting an LCA (this standard was last reviewed and confirmed in 2022; therefore this version remains current). ISO 14040 specifies four main phases of an LCA: goal and scope definition, inventory analysis, impact assessment, and interpretation. The inventory analysis phase, which is the stage where the inputs and outputs of a product's life cycle are quantified, typically includes the following:

- Inputs: These are the materials, energy, and water that are required for a product or process to be produced, transported, and used. Inputs can include raw materials, water, energy sources such as fossil fuels and electricity, and other consumables like chemicals or packaging materials.
- Outputs: These are the emissions, waste, and other environmental impacts that result from the production, transportation, and use of a product or process. Outputs can include air emissions like greenhouse gases, particulate matter, and volatile organic compounds (VOCs), as well as water and soil pollution, waste generation, and other environmental impacts.

9. While there is some flexibility in the specific inputs and outputs that are included in an LCA, it is generally recommended that LCAs be conducted using a comprehensive set of data that captures the full life cycle of a product or process. This helps to ensure that the LCA provides a complete and accurate picture of the product's environmental impacts.

10. The normalised impact values used in LCA studies represent the relative magnitude of different environmental impacts associated with a product or process. The specific size of the difference between normalised impact values

depends on the specific methodology and characterisation factors used in the LCA study. ISO 14074 (published in November 2022) sets out guidelines, requirements, and principles for normalisation, weighting, and interpretation of LCAs.

11. Normalisation is a process in which the environmental impact categories that were identified and quantified in the LCA study are aggregated and compared to reference values. This allows the environmental impacts of a product or process to be expressed as a ratio or percentage of a reference value, which can be used to compare the relative magnitude of different impacts. For example, the Global Warming Potential (GWP) impact category is commonly used in LCAs to evaluate the potential contribution of greenhouse gas emissions to climate change. GWP is expressed as a normalised impact value in units of CO<sub>2</sub> equivalents (CO<sub>2</sub>eq), where the impact of different greenhouse gases, such as methane and carbon dioxide, are multiplied by their respective Global Warming Potential factors to convert them to an equivalent amount of CO<sub>2</sub>. The difference in impact between two normalised GWP values would depend on the specific values used for the GWP factors.

12. Normalised impact values are relative and not absolute measures of environmental impact. The absolute magnitude of a given impact will depend on a variety of factors, including the quantity of emissions or other releases, the location and timing of the emissions, and the specific environmental conditions in which they occur. However, normalised impact values can provide a useful tool for comparing the relative environmental performance of different products or processes within a given impact category.

13. There are various regulatory guidance documents for conducting LCAs across different countries and regions around the world. These documents provide guidance on the methods, data sources, and reporting requirements for conducting LCAs in a way that meets the needs of regulatory agencies.

14. In the European Union (EU), for example, the European Commission has developed a set of guidelines for the environmental assessment of products, which includes requirements for conducting LCAs. The guidelines, known as the [Product Environmental Footprint \(PEF\)](#), provide a standardised methodology and data requirements for conducting LCAs of products.

15. In the United States, the Environmental Protection Agency (US EPA) has developed [guidance](#) for conducting LCAs in a range of industries and sectors, including agriculture, building materials, electronics, and transportation. The

guidance documents provide information on data sources, methods, and reporting requirements for conducting LCAs that meet the needs of regulatory agencies.

16. Other countries and regions, including Canada, Japan, and China, have also developed regulatory guidance for conducting LCAs. These documents often draw on international standards and guidelines, such as the ISO 14040 series of standards for LCA, to provide a common framework for conducting LCAs that can be used by regulatory agencies and other stakeholders.

## **Questions for the Committee**

- 1) Do Members have any comments on the new information included in the position paper (as outlined in paragraph 4 above)?
- 2) Do Members have any comments on the information on life cycle analysis?
- 3) Would the Committee like the information on LCA to be included in the position paper, and if so would a brief synopsis be sufficient?
- 4) Does the Committee have any comments on the structure or content of the draft position paper?
  - i. Does the position paper (annex A) sufficiently outline and summarise the discussions thus far on allergenicity of chitin and chitosan based BBFCMs?
- 5) Does the Committee have any recommendations for future research regarding chitosan in BBFCMs?

### **Secretariat**

**September 2023**

## **Annex A to TOX/2023/43**

### **Background**

1. The use of fossil-based plastics has been associated with adverse environmental impacts. Consequently, there is interest in reducing the amount of conventional plastic used for packaging, and recent years have seen a major global increase in the development and use of bio-based food contact materials

(BBFCMs). Bio-based materials are defined as being derived, directly or indirectly, from a renewable source of living matter (Bradley, 2010).

2. Some BBFCMs under development contain chitosan, which is a biodegradable polysaccharide derived from chitin (Figure 1). Chitosan has antimicrobial and antioxidant properties which make it ideal for extending the shelf-life of packaged foods (Vasile, 2018).

3. Chitin is a high molecular weight  $\beta(1,4)$ -linked homopolymer of N-acetylglucosamine (see Figure 1). In situ, chitin is linked to other structural components, such as protein and glucan, to form a protein-chitin matrix (Romano et al., 2007). Chitin is converted to chitosan by removing the acetyl groups (COCH<sub>3</sub>).

Figure 1: Chemical structures of chitin (R<sub>1</sub> = COCH<sub>3</sub>) and chitosan (R<sub>1</sub> = H).

4. Presently, the main commercial source of chitosan is from chitin obtained from waste streams of the fishing industry, i.e. crustacean shells. However, the recent increased global demand for chitosan has drawn attention to other possible sources: fungi and insects.

5. Production of chitosan from chitin involves deproteination and subsequently deacetylation. However, since the level of deproteination reported in studies from the literature is 100 % and any residual protein in chitosan might contain allergenic proteins (Yadav et al. 2019), there is a concern regarding the potential allergenicity of chitosan when used in FCM.

## Shellfish allergy

6. According to [Anaphylaxis UK](#) (a charity supporting people living with serious allergies), there are different types of shellfish which can all cause allergic reactions. Shellfish can be split up into two groups, crustaceans and molluscs. Crustaceans include crab, lobster, crayfish and prawns. Molluscs can be split up further into bivalves (which include mussels, oysters, razor shells, scallops and clams), gastropods (which include limpets, periwinkles and snails found on land) and cephalopods (which include squid, octopus and cuttlefish).

7. Anaphylaxis UK state that “If you react to one type of shellfish it’s likely you’ll react to others in the same group. For example, if you

react to crabs, it's likely you'll react to other crustaceans. You might react to shellfish in the other group as well, in this case molluscs". This phenomenon is called cross-reactivity.

8. **Anaphylaxis UK go on to explain that, "An allergic reaction (including anaphylaxis) happens when the body's immune system wrongly identifies a food or substance as a threat. When this happens, the body releases chemicals, such as histamine, in response. It is the release of these chemicals that causes symptoms". Furthermore, "The symptoms of a shellfish allergy usually come on quickly, within minutes of eating the food. Mild to moderate symptoms may include: a red raised rash (known as hives or urticaria) anywhere on the body, a tingling or itchy feeling in the mouth, swelling of lips, face or eyes, and stomach pain or vomiting. The term for a more serious reaction is anaphylaxis. Most healthcare professionals consider an allergic reaction to be anaphylaxis when it involves difficulty breathing or affects the heart rhythm or blood pressure". Anaphylaxis can be life-threatening.**

9. **In their review of shellfish allergy, Lopata et al. (2010) noted that cross-reactivity occurs frequently to seafoods within a certain group or family such as crab, lobster, shrimp among the crustaceans, suggesting that cross-reactivity frequently occurs between phylogenetically related organisms. Furthermore, crustacean allergic subjects often react to species of the mollusc group, such as squid (cuttlefish), abalone, limpet, squid, oyster, mussel, scallop and clam. Lopata et al. concluded that tropomyosin seems to be the major allergen responsible for molecular and clinical cross-reactivity between crustaceans and molluscs.**

10. **The major allergen identified in crustaceans is tropomyosin, although several other allergenic proteins including arginine kinase and myosin light chain have been identified in shrimp and other crustacean shellfish as well as molluscan shellfish (EFSA, 2014).** Tropomyosin is a muscle protein present in all species of vertebrates and invertebrates. However, only the tropomyosin found in invertebrates such as crustaceans, arachnids, insects, and molluscs has been associated with allergic reactions in humans (Reese et al., 1999).

11. Tropomyosin is a heat-stable allergen (Daul et al., 1994). It is also an "acidic" protein with an isoelectric point (pI) value of 4.5 (Reese et al., 1999). Due to these characteristics, tropomyosin can be present in processed foods (Reese et

al., 1999). However, in their review of shellfish allergy, Woo & Bahna (2011) note that tropomyosin's "allergenicity may change by certain processing methods. Boiling may result in the Maillard reaction (glycation) and formation of neoepitopes, as demonstrated that in some patients, boiled shrimp extract induced larger skin test responses than raw extract. Also, shrimp extract treated with high intensity ultrasound for 180 minutes demonstrated decreased binding with sera from shrimp allergic patients".

12. In the PhD thesis of Nguyen (2012), results of a dot blot experiment demonstrated the presence of tropomyosin in technical samples of chitin and chitosan samples (these samples were provided by Mahidol University in Thailand). Furthermore, an inhibition enzyme-linked immunosorbent assay (ELISA) confirmed the immunoreaction of the protein residue in chitin and chitosan samples to rabbit sera (which was specific to shrimp tropomyosin). Subsequently, Nguyen (2012) noted that "special care should be taken when using chitin and chitosan in food or medical preparations. Warning statements should state clearly the presence of tropomyosin in products derived from chitin or chitosan, especially when the consumers are sensitised to crustaceans".

## **Chitin & chitosan-based BBFCMs in development**

13. Food packaging materials containing chitosan are being developed in the form of flexible films or coatings. A "film" is preformed separately and wrapped onto a food surface. These films are usually prepared by using a solvent casting method, in which chitosan is dissolved in suitable solvents (in most cases, slightly acidified water) and then poured onto a flat surface to allow the solvent to evaporate (Kim et al., 2006). On the other hand, a "coating" is a thin layer formed directly onto the surface of food **and/or its packaging**. Direct application of chitosan formulations onto food surfaces can be attained by spraying or dipping (Tharanathan, 2003). Additionally, drinking straws which contain chitosan have also been developed (paragraph 39).

14. In studies found in the literature, chitosan-based films may be described as edible or inedible, whereas chitosan coatings are almost always described as edible since they form a layer directly on the top surface of the food (Priyadarshi & Rhim, 2020). Another difference is that chitosan films are  $>30\ \mu\text{m}$  in thickness, whilst coatings are  $30\ \mu\text{m}$  in thickness (Van den Broek et al., 2015). However, with the advancement of nanotechnology, nano-coatings are being explored, which consist of nanoscale layers (100 nm) built-up onto food surfaces (Vasile, 2018). In their review of nanoedible films for food packaging, Jeevahan &



Chandrasekaran (2019) noted that production of edible films and coatings was still largely at the laboratory level and was not yet expanded to industrial level due to their high cost of production.

15. It has been suggested that chitosan-based films could appear in vacuum-packaged processed meat (Ouattara et al., 2000), cheese (Fajardo et al., 2010), and other foods such as vegetables, fruits, grains, and fish (Sinha et al., 2012).

16. Modifying chitosan by the addition of a metal has been shown to enhance its antimicrobial activity compared to native chitosan (Du et al., 2009). Subsequently, some chitosan-based BBFCMs in development are nanoengineered to contain metal ions, such as copper (Yin et al., 2018).

17. Some BBFCMs in development contain chitin, in the form of nanofibers (Ifuku & Saimoto, 2012) or nano-whiskers (Zeng et al., 2012). Incorporation of chitin nano-whiskers into starch-based films has been shown to improve the film's mechanical and barrier properties (Qin et al., 2016). For chitin nano-whiskers migration studies are scarce, mainly due to the difficulties in characterising nanoparticles in composites generally, and the lack of methods for qualitative and quantitative analysis (Han et al., 2011).

## **Market uses of chitosan**

18. Chitosan has applications in tissue engineering and biomedicine due to its low cost, biocompatibility, lack of toxicity, and biodegradability (Madhumathi et al., 2009; Konovalova et al., 2017).

19. Chitosan is widely used as a food additive and functional ingredient in foods sold in Italy, Finland, Korea and Japan (Peter, 1997; Singla & Chawla, 2001). Both chitin and chitosan are approved food additives in Japan (JFCRF, 2011). Furthermore, chitosan is listed as a processing aid in the Codex General Standard for Fruit Juices and Nectars (Codex, 2005). The Norwegian company "Norwegian Chitosan AS" trades chitosan (Kitoflokk™ and Norlife) for several applications, including food and beverages (Ferreira et al. 2016).

20. Chitosan and chitin have not been officially classified by the US Food and Drug Administration (US FDA) but some biotechnology companies have notified the US FDA of their view that the use of chitosan and chitin in specific food applications would be generally recognised as safe (GRAS). For example, KitoZyme views the use of chitosan (derived from *Aspergillus niger*) in alcoholic beverage production (with chitosan being removed from the beverages post-

treatment, using physical separation processes) as GRAS. In their correspondence to KitoZyme, the US FDA (2011) wrote: “based on the information provided by KitoZyme, as well as other information available to FDA, the agency has no questions at this time regarding KitoZyme’s conclusion that chitosan from *A. niger* is GRAS under the intended conditions of use. The agency has not, however, made its own determination regarding the GRAS status of the subject use of chitosan”.

21. Shellfish-derived chitosan is sold online as a dietary supplement, with manufacturer-recommended daily consumption of chitosan, for example, of [2.4 g](#) and [3 g](#). It has been suggested that chitosan may support weight loss and lower cholesterol by eliminating fat and cholesterol from the body instead of allowing the body to absorb them (Moraru et al. 2018).

22. Chitosan is considered to be hemostatic due to its cationic nature (NTP, 2017), which supports its use in wound dressings. Wound dressings manufactured from shellfish-derived chitosan were introduced in 2005 for US soldiers, and in 2008 the US FDA approved the HemCon bandage for use as a dressing for local management of bleeding wounds (US FDA, 2008).

## **Absorption, distribution, metabolism, and excretion (ADME)**

23. Results from Chae et al. (2005) indicated that absorption of chitosan from the gastrointestinal (GI) tract following oral exposure in rats was inversely related to its molecular weight: oral gavage administration of chitosan with molecular weights of 3.8, 7.5, 13, 22, or 230 kDa resulted in maximum plasma concentrations of 20, 9, 6, 4, or 0.5 µg chitosan/mL, respectively.

24. Several chitinases have been identified in humans which can bind and degrade chitin (Boot et al., 2001). Furthermore, *Lactococcus lactis* and *Lactobacillus plantarum* have chitinolytic and/or chitin-binding proteins (Sánchez et al., 2011). These bacteria are an integral part of normal gut flora, fermented foods, and probiotic-fortified foods (Kim et al., 2013; Todorov et al., 2012).

25. Degradation of chitosan in vertebrates is thought to occur predominantly by lysozymes and bacterial enzymes in the colon (Kean & Thanou, 2010).

26. The depolymerised products of chitin or chitosan are called chitooligosaccharides (COS), which have a molecular weight of approximately **3.9**

**kDa or less (Lodhi et al., 2014).** COS are water-soluble (Qin et al., 2006), and have antioxidative, anti-inflammatory, and antibacterial effects (Huang et al., 2016). However, COS have been observed to irritate intestinal epithelial mucosal tissues, stimulating them to produce mucin (Deters et al., 2008). Following depolymerisation, both chitin and chitosan particles are readily phagocytosed (Bueter et al., 2011).

## Allergen reference doses

27. The major allergen identified in crustaceans is tropomyosin. However, no threshold levels for tropomyosin alone could be identified in the literature. However, studies conducted by Remington et al. (2020) and Houben et al. (2020) determined allergen eliciting doses for crustacean-derived protein, i.e. an ED01 (where 1 % of the allergic population may be expected to react) of 25 mg (95 % confidence interval of 2.7-166 mg) for shrimp protein, and an ED05 of 280 mg (95 % confidence interval of 69.3-880 mg). The ED01 and ED05 were derived from human food challenge data using dose-distribution modelling and represent acute intake levels of crustacean-derived protein that are predicted to provoke any objective reaction in no more than 1 and 5 % (respectively) of at-risk individuals, who show a minimal allergic response upon challenge. However, the severity of these reactions is variable. The large confidence intervals for both EDs reflect the uncertainties around those estimates for this allergenic food.

**28. In 2023, an Expert Committee of the Food and Agriculture Organization of the United Nations (FAO) and the World Health Organization (WHO) published a meeting report on the risk assessment of food allergens, which included consideration of threshold levels in foods for eight priority allergens (FAO/WHO, 2023). The Expert Committee recommended that reference doses (RfDs) should be based on ED05 values (FAO/WHO, 2023). The shrimp ED05 value of 280 mg shrimp protein was rounded down to a single significant figure on the basis of the size of the confidence interval. Thus, the Expert Committee recommended a RfD for crustacea of 200 mg crustacea (shrimp) protein because the analysis relied on a few species of shrimp to provide data for the group of crustacea and the additional, more conservative rounding was considered appropriate when considering the diversity of crustacea species consumed (FAO/WHO, 2023).**

29. **A subgroup of the Committee on Toxicity of Chemicals in Food, Consumer Products and the Environment (COT) assessed the Joint**

**FAO/WHO Expert Committee's report to inform the UK Food Standards Agency (FSA) and Food Standards Scotland (FSS) risk managers' understanding of whether it is appropriate for the recommended reference doses based on ED05 values to be applied to the UK 14 regulated allergens, including crustaceans ([TOX/2023/35](#)). In addressing the questions posed in the Terms of Reference, the COT subgroup reached the following conclusions:**

- a) The COT subgroup were of the view that there is no reason to suppose the data on which the Codex Expert Committee based their analysis are not sufficiently representative of the UK population.**
- b) Key gaps identified by the COT subgroup that need to be addressed before the UK can adopt the recommended RfDs included some uncertainties regarding the way in which the ED values have been derived. However, the key gaps identified could not be filled using the published literature.**
- c) There is insufficient evidence to demonstrate that using RfDs based on ED05 instead of ED01 values would not significantly impact on public health.**

**30. The COT subgroup's draft report was discussed by the full Committee at the July 2023 COT meeting. Members agreed with the COT subgroup's methodology of assessment and the contents of the draft report, including the conclusions. Work by the FSA/FSS to determine appropriate allergen thresholds is still ongoing.**

31. In EFSA's 2014 evaluation of allergenic foods and food ingredients for labelling purposes, EFSA noted that "studies reporting on the prevalence of allergy to crustaceans in the general (unselected) European population are scarce. In the few studies available, the prevalence of self-reported crustacean-related adverse reactions to food in children ranged from 0.1 % and 0.3 % in Greece (Zannikos et al., 2008) and the UK (Pereira et al., 2005) to 5.5 % in France (Touraine et al., 2002). Figures reported from the Netherlands (Brugman et al., 1998), Sweden and Iceland (Kristjansson et al., 1999) were within that range (0.7–1.5 %). Prevalence of self-reported allergy to shrimp was 0.5 % in 2- to 14-year-old Finch [sic] children (Rancé et al., 2005). In adults, estimated sensitisation rates to crab in Germany (Schafer et al., 2001) based on positive skin prick tests (SPT) were similar to those reported in Hungary (Bakos et al., 2006) based on specific immunoglobulin E (IgE) testing (1.9 % and 1.8 %, respectively).

Prevalence rates of allergy to crustaceans based on clinical history and positive SPT in the German general population were much lower (0.2 %) (Zuberbier et al., 2004). Only one study conducted in Denmark reported challenge proven prevalence data for shrimp allergy, which ranged from zero in subjects 22 years to 0.3 % in subjects > 22 years (Osterballe et al., 2005)”.

32. Pereira et al. (2005) investigated the rates of food hypersensitivity (FHS) in UK teenagers against a panel of allergens including shellfish. These teenagers (11 and 15 year-olds) were resident on the Isle of Wight at the time of the study, and completed a questionnaire with their parents on adverse reactions to foods. Results showed that two 11 year-olds (0.3 %; n = 699), and five 15 year-olds (0.8 %; n = 649) reported an adverse event from consumption of shellfish.

33. From an analysis of UK hospital admission data from 1998 to 2018, Conrado et al. (2021) concluded that "hospital admissions for food induced anaphylaxis have increased from 1998 to 2018, however the case fatality rate has decreased". From 1998 to 2018, in children (16 years old) fish & crustacea triggered 6 % of all cases of fatal food induced anaphylaxis, whereas for adults it was 7 %.

## **Case reports of reactions to chitosan**

34. Kato et al. (2005) reported a case of immediate-type allergy from use of a health food containing chitosan, where “the patient was a 47-year-old female person who developed systemic urticaria and difficulty in breathing after oral ingestion of chitosan. Since skin tests (prick test and scratch patch test) were positive, the test was done using another commercial chitosan, and was positive. The patient was diagnosed as having chitosan-induced immediately-type allergy, and was instructed to avoid ingestion of chitosan. The patient developed no symptoms thereafter”. The study authors concluded that chitosan may have functioned as a food allergen because of its molecular weight and general properties.

35. Two case reports were identified relating to hypersensitivity to some healthcare products containing chitosan (Cleenewerck et al., 1994; Pereira et al., 1998). The biological source of the chitosan was not stated in these publications.

## **Reactions from entomophagy**

36. Reports on adverse reactions from insect consumption (entomophagy) are scarce and only two population studies (described below) were identified in

the literature which reported on the prevalence of food allergy to insects. In these two studies, clinical measurements of allergy do not seem to have been verified, which was a limitation of the data.

37. Taylor & Wang (2018) investigated the prevalence of allergic reactions caused by consuming edible insects. The investigation was conducted in the North Eastern (or the Isan region) of Thailand, in an area where entomophagy is common. Information concerning entomophagy and allergic reactions were gathered from multiple sources in four locations: Nongki, Nang Rong, Nong Bun Mak, and Nakhon Ratchasima. The survey included questions about eating habits in relation to insects, other known food allergies, and presented a list of symptoms the participants may have experienced. The prevalence of allergic reactions caused by consuming edible insects was much higher than expected across the 2,500 respondents. In the Isan region, approximately 14.7 % of people experienced a single symptom indicative of an edible-insect allergy, and 7.4 % of people experienced multiple symptoms indicative of an edible-insect allergy. Furthermore, approximately 46.2 % of people that already suffered from a known food-based allergy also experienced symptoms indicative of an allergic reaction after insect consumption. According to the study authors, “the most common symptoms appear to be gastrointestinal (diarrhoea and vomiting)”. The study authors concluded that “the allergy aspect of entomophagy is a serious issue and has the potential to adversely affect the future of entomophagy, especially in introducing the concept to western cultures”.

38. Barennes et al. (2015) assessed the prevalence of food allergy to insects amongst insect-eaters. In this survey, eight teams (which included medical physicians) collected data to address socioeconomic characteristics of the consumers, types of insects consumed, frequency of consumption and reports of side effects. This study was conducted in Laos, and included 1,059 subjects that had previously eaten insects, 81 of whom (7.6 %) reported “allergy problems after eating insects”. Of these 81 subjects, 38 reported that allergy problems were “mostly with grasshoppers or stink bugs”. None of the subjects reported severe anaphylaxis. In this survey, it was not possible to identify how much the consumption of edible insects contributed to the daily diet of the population or provide detail on the way insects were harvested. The survey did not mention whether any clinical confirmation of the allergenic symptoms was undertaken.

### **UK incidents**

39. The FSA has received a number of queries about the presence of chitosan in food packaging materials and chitosan-based drinking straws, but no incidents have formally been raised within the FSA. However, there was one report of a potential allergic reaction to the use of a chitosan-based straw in a pub which was reported to a local authority. The local authority carried out an investigation with the supplier of the chitosan-based straws but it was difficult to rule out cross-contamination from the meal that the individual had also consumed on the premises. The individual who suffered the allergic reaction did have a seafood allergy but did not disclose this to the pub. That was the only incident reported to the FSA but uncertainties remain over the cause of the allergic reaction.

## UK legislative position

40. In retained European legislation, all materials and articles intended for contact with food must meet the requirements of the Framework Regulation (EC) [No. 1935/2004](#). The principle underlying this Regulation is detailed in Article 3 which states: “materials and articles, including active and intelligent materials and articles, shall be manufactured in compliance with good manufacturing practice so that, under normal or foreseeable conditions of use, they do not transfer their constituents to food in quantities which could: a) endanger human health; b) bring about an unacceptable change in the composition of the food; c) bring about a deterioration in the organoleptic characteristics thereof.”

41. With regards to necessary labelling (and potential exposure to allergens) Article 15 of retained Regulation (EC) [No 1935/2004](#) states that ‘special instructions (are) to be observed for safe and appropriate use’. This labelling information may need to be provided on the packaging, or as a standalone warning should the item be sold loosely. If the item is marketed as edible, other labelling requirements come into play to comply with food law and the Materials and Articles in Contact with Food Regulations 2012 as amended.

42. Whilst there are no specific migration limits for BBFCMs, industry can refer to legislation that may be pertinent (the same holds true for other materials lacking specific legislation). **Retained Regulation (EU) [No 10/2011](#) states that: “According to good manufacturing practice it is feasible to manufacture plastic materials in such a way that they are not releasing more than 10 mg of substances per 1 dm<sup>2</sup> of surface area of the plastic material”**. Therefore, the Plastics Regulation stipulates a generic migration limit (maximum allowable) of 10 mg per square decimetre of surface area of material (10 mg/dm<sup>2</sup>). **According to Retained Regulation (EU) [No 10/2011](#), “The**

**overall migration limit of 10 mg per 1 dm<sup>2</sup> results for a cubic packaging containing 1kg of food to a migration of 60 mg per kg food. For small packaging where the surface to volume ratio is higher the resulting migration into food is higher”**. The applicability of FCM legislation depends on the BBFCM’s intended use and how it is marketed. If the BBFCM is intended purely for containment purposes and is inedible, it is not food and comes under FCM legislation.

43. The EU considers that an edible film is a special active part of the food and, seen from a legal point of view, it is to be regarded as a foodstuff, along with the food packed in the film, having to fulfil the general requirements for food (Fabec et al., 2000); this is also the position of the FSA FCM Policy team. Subsequently, the presence of a known allergen in an edible film or coating on a food must be clearly stated on the label (Campos et al., 2011). Due to hygienic reasons, it is anticipated that food products in edible films need to have an outer package, otherwise the film should not be eaten (Fabec et al., 2000).

## **Evaluations of crustacean chitosan**

### **2011 Evaluation by EFSA (EFSA NDA Panel)**

44. In 2011, when reviewing a proposed health claim for a food supplement containing crustacean-derived chitosan, the EFSA Dietetic Products, Nutrition and Allergies (NDA) Panel concluded, that “a cause and effect relationship has been established between the consumption of chitosan and maintenance of normal blood LDL-cholesterol concentrations”, and “considers that in order to obtain this effect in adults, 3 g of chitosan should be consumed daily” (EFSA, 2011). The Panel stated that their opinion does not constitute, and cannot be construed as, a positive assessment of its safety.

### **Studies from the literature**

45. Studies designed to evaluate the effectiveness of shellfish-derived chitosan as an oral weight-loss supplement over 12 days suggest that it is well tolerated in men and women at 4.5 g chitosan per day (Gades & Stern 2003, 2005). However, these studies excluded subjects with food allergies and sensitivities. Data collection sheets for the volunteers did not appear to have a space for recording any adverse effects, but one of the 15 male participants reported “vomiting after a meal during the supplement period” (Gades & Stern 2003). Additionally, in a study involving 65 men and women, consumption of



chitosan tablets (6.75 g of chitosan daily for eight weeks), was “found to be safe”, though common transient GI symptoms were reported (loose faeces, constipation, abdominal pain, repeated flatulence, abdominal bloating, and abdominal rumbling) (Tapola et al., 2008). However, the study excluded subjects with a history of severe allergic reactions (anaphylactic reaction) when exposed to fish or crustacean.

46. In 2011, Waibel et al. (2011) investigated the safety of “HemCon®” bandages in patients who reported a shellfish allergy. Initial assessment included a detailed history, IgE SPT, and serum testing to shellfish allergens. Participants who demonstrated specific shellfish IgE underwent a bandage challenge. It was reported that of the 19 participants who were enrolled, ten completed the study **as they had met the inclusion criteria**. Seven (70 %) were male and the average age was 44.8 + [sic] 10 years. Nine (90 %) reported a shrimp allergy history and five (50 %) reported multiple shellfish allergies. All participants completing the study had positive SPT and serum IgE testing to at least one shellfish; eight (80 %) had shrimp positive SPT and ten (100 %) demonstrated shrimp-specific IgE. No participant had a positive SPT to chitosan powder or experienced an adverse reaction during bandage challenges. No protein bands were visualised during gel electrophoresis analysis of chitosan powder. The study authors concluded that all participants tolerated the HemCon bandage without reaction.

## Evaluations of fungal chitin

### 2010 Evaluation by EFSA (EFSA NDA Panel)

47. In 2010, the EFSA panel on NDA assessed the safety of chitin-glucan as a novel food ingredient (EFSA, 2010). This chitin-glucan was derived from *A. niger* through a fermentation process, and therefore did not contain shellfish protein. The product (KiOnutrime-CG™) assessed by EFSA was composed of >90 % chitin-glucan (a structure that combines chitin and beta (1,3) glucan) and ≤ 6 % protein, and was intended to provide a daily intake of 2 - 5 grams of chitin-glucan. The Panel reviewed a report showing no observed adverse effects at the highest dose administered (about 6.6 g/kg bw) in a 13-week rat study (TNO, 2009). Because this dose is approximately 80-fold higher than the maximum intended level of intake for humans on a g/kg bw basis, the Panel concluded that KiOnutrime-CG™ was safe as a food ingredient at the proposed conditions of use and at the proposed intake levels. The Panel assessed the risk of allergenicity on the basis of

some allergenic enzymes that are synthesised by *A. niger* such as beta-xylosidase. The Panel concluded that “an allergenic risk cannot be ruled out but is expected not to be higher than from the consumption of other *A. niger* derived products”. The Panel also noted that since *A. niger* is commonly detected in various foods such as fruits and vegetables, it is therefore expected to occur in the diet of most individuals.

## **2012 Evaluation by FSANZ**

48. In 2012, Food Standards Australia New Zealand (FSANZ) approved an application for the use of *A. niger*-derived chitosan as a processing aid for production of some beverages. In their risk assessment, FSANZ noted that animal toxicity studies on chitosan preparations of various molecular weights and degrees of acetylation did not show any treatment-related adverse effects following oral administration at high doses. Furthermore, “a published review of human data from 13 clinical trials of up to six months duration found no adverse effects associated with oral chitosan (average daily dose 3.5 g) as a weight loss supplement. In view of the absence of adverse effects at high chitosan doses, a group acceptable daily intake (ADI) “not specified” was established for chitosan derived from fungi. Information was provided indicating negligible levels of fungal chitosan in wine following processing. Negligible levels would also be expected in beer and cider, while no residual fungal chitosan would be expected in alcoholic products derived from distillation”. The overall conclusion was that the “use of fungal chitosan as a processing aid for the production of wine, beer, cider, spirits and food grade ethanol is technologically justified and raises no public health and safety issues for consumers” (FSANZ, 2012).

### **Studies from the literature**

49. Seaton & Wales (1994) conducted an 8-year follow-up study on clinical reactions to *A. niger* in a biotechnology plant producing citric acid by fermentation of molasses with *A. niger*. The authors concluded that *A. niger* was a weak antigen, and that simple hygiene measures were needed to protect the workforce.

50. Mycoprotein is a food produced for human consumption by fermentation of *Fusarium graminearum* on a glucose substrate. Known as ‘Quorn’, it is widely available in most leading UK retailers where it has been on sale since 1985. To investigate whether mycoprotein would cause allergic responses either in exposed production workers, or in those ingesting Quorn, immunological studies

were conducted by Tee et al. (1993). Mycoprotein production workers were screened for allergy using the radioallergosorbent test (RAST) over a 2-year period. Two of ten patients referred to hospital following vomiting and diarrhoea after ingestion of mycoprotein had a mycoprotein skin-prick test (SPT) response  $\geq 2$  mm, but none had a significantly raised RAST. Specific IgE antibody to mycoprotein was not significantly raised in any complainant. The study authors concluded that “the possibility cannot be excluded of participation of fungal polysaccharide allergens, which RAST testing would probably not detect, or of non-IgE associated mechanisms. Intolerance to ingested Quorn reported by a small number of consumers may be due mainly to an idiosyncratic response”.

## Evaluations of insect chitin

### 2020 Evaluation by EFSA (EFSA NDA Panel)

51. In 2020, the EFSA NDA Panel published their opinion on the safety of dried yellow mealworm (*Tenebrio molitor* larva) as a novel food (NF) (EFSA, 2020). **This evaluation was on the whole mealworm preparation (not just the insect chitin), and therefore was a more holistic assessment of a potential allergic response.** The reported average chitin content of the NF in powder form was reported to be  $6.42 \pm 0.28$  g/100 g across five batches (the NF was not reported to contain chitosan).

52. The NDA Panel noted that “yellow mealworms are consumed as part of the customary diet or for medicinal purposes in some non-EU countries worldwide. Their consumption by humans has been reported in Thailand (Hanboonsong et al., 2013), China (Feng et al., 2018) and Mexico (Ramos-Elorduy, 1997, 2009; Ramos-Elorduy and Moreno, 2004). Yellow mealworms are among the insect species permitted to be consumed as food in Korea by the Korean Food and Drug Administration (KFDA) (Kim et al., 2017). Additionally, in Australia and New Zealand yellow mealworms are considered as non-traditional, not novel foodstuff (FSANZ, 2020). Since 1 May 2017, *T. molitor* larva is among the insect species that can be legally introduced in the Swiss market as food (whole, chopped or ground)” (EFSA, 2020). Because of this history of use, and the absence of adverse effects described in the literature, the Panel concluded that “the NF is safe under the proposed uses and use levels”. The proposed use was as an ingredient in several food products, such as pasta-based dishes, and biscuits, for all population groups.

53. The NDA Panel also discussed the work of Broekman et al. (2017) which demonstrated the possibility of de novo human sensitisation to allergens in mealworm, which can result in food allergy. Broekman et al. (2015) demonstrated that thermal processing did not lower the allergic potential of mealworm allergens (EFSA, 2020). The Panel also noted that “the applicant provided the study of Velasquez (2015) who investigated the allergenic potential of yellow mealworm larvae using extracts of the NF and concluded that subjects allergic to arthropods and more specifically to crustaceans, should not consume the NF due to the risk of cross-reactivity”. Subsequently, the NDA Panel considered that “the consumption of the NF may induce primary sensitisation and allergic reactions to yellow mealworm proteins and may cause allergic reactions in subjects with allergy to crustaceans and dust mites. Additionally, allergens from the feed may end up in the NF”. Furthermore, the Panel recommended that research should be undertaken on the allergenicity of yellow mealworm, including cross-reactivity to other allergens.

### **Studies from the literature**

54. Broekman et al. (2016) included 15 patients with shrimp allergy (based on specialist opinion and diagnostic testing) in a double-blind, placebo-controlled food challenge (DBPCFC) trial, and found that 13 of these patients also had mealworm allergy. The study authors noted that “when comparing the mealworm challenge outcome of 4 patients who also had a shrimp challenge, eliciting doses (ED5 and ED10) as well as severity were in the same range”, which “indicate(s) that mealworm is at least as allergenic as shrimp”, though “more mealworm challenge data are needed to confirm this initial analysis”. Subsequently, Garino et al. (2020) used the data from Broekman et al. (2016) to predict values for the ED05, ED10, and ED20, indicating where 5 %, 10 % and 20 % of the shrimp allergic population are predicted to react to mealworm proteins. The values for the ED05 were all 63, 128, and 147 mg of *T. molitor* protein, estimated using the Weibull, log-logistic, and log-probit distribution models, respectively.

### **FSA response**

55. In addition to seeking the advice of the COT, the FSA is corresponding with a biotechnology company which is developing food packaging materials comprised of chitosan, as this provides a useful addition to the current state of knowledge for these materials.

## COT consideration

56. The COT considered that additional information was needed in order to assess the risk of allergenicity of chitin- or chitosan-based BBFCMs. More information would be needed, in particular on the effects of processing but also additional data characterising the protein content in chitosan and the final BBFCMs (against chemical and enzymatic methods of deproteination), data on migration from, and consumption of, BBFCMs. Information on the total amount of residual protein (expressed as mg/g BBFCM) would be helpful for estimating health risks.

57. In order to assess the health risk from the use of BBFCMs containing chitosan, additional information would be needed, notably on the level of any allergenic proteins migrating from these packaging materials into food. Although chitosan has been incorporated into the composition of some BBFCM products, it was not clear whether the manufacturing processes of these BBFCMs further reduced the protein content within the chitosan and/or the allergenicity of these proteins. Therefore, it would be helpful to obtain protein migration data from BBFCM products that contain chitosan under expected conditions of use, alongside estimates of consumer use/consumption of these products in order to provide an estimation of consumer exposure to any allergenic proteins.

58. The COT noted that available clinical ingestion data (Gades & Stern 2003, 2005; Tapola et al., 2008) indicated that the immunological properties of chitin and chitosan were of low concern in the context of BBFCMs. Chitin was well tolerated in supplements at higher exposures than would be expected from the use of BBFCMs. However, some adverse effects were associated with high intakes of the raw materials in clinical studies, which were typically mild symptoms of gastrointestinal tract distress such as diarrhoea, bloating, or vomiting. The indications were however, that this was a relatively non-specific inflammatory reaction. The COT agreed that these adverse effects were not of concern for BBFCMs as the processing was likely to produce a more inert final material. Furthermore, the phagocytosis of small fragments of chitin or chitosan appeared to be the same as that of similar-sized particles in general.

59. The COT agreed that the limited information provided in this case report of immediate-type allergy for chitosan-containing health food (Kato et al., 2005) did not suggest any additional concerns. The COT noted that this reported case of immediate-type allergy was most likely due to residual protein from the shellfish source from which the chitosan supplement was derived. The COT agreed that the

type of hypersensitivity described in the two case reports of hypersensitivity to some healthcare products containing chitosan (Cleenerwerck et al., 1994; Pereira et al., 1998) very rarely, if ever, occurs in the context of food ingestion.

60. A submission by Primex to the US FDA in 2012 (GRAS Notice No. 443) contained a dossier including some approaches to protein measurement and analytical data for the ED01 and corresponding analysis. The COT noted that the chitosan used in this submission appeared to be highly controlled in terms of its production, and whilst its specification may be unlike that of other chitosan products, it nevertheless provided a standard to be achieved and possibly put forward. The Committee agreed that during its manufacture, chitosan needed to have a certain specification in terms of protein content.

61. The COT considered that the ED01 was an adequate protection goal, given the potential for increased human exposure to the allergen if it were to be present in food packaging. However, work is still underway with the COT thresholds sub-group to inform FSA decisions on which ED should be used. It was agreed that the choice of benchmark (e.g. ED01) was a risk management decision. Due to the large amount of data required for dose distribution modelling, accurate estimates below ED01 were not feasible.

62. The COT considered that in order to assess whether FCM posed a negligible health risk in practice (if consumption was below the ED01), it would be necessary to understand the effects of processing on the levels of allergens in the final FCM, and the extent to which they may migrate into food (as is the case for other allergens).

## **Next steps**

63. At present, no measurements of the amount of allergenic protein in BBFCMs have been identified in the scientific literature. Furthermore, no public usage or consumption data for chitin or chitosan based BBFCMs were identified in the literature or the National Diet and Nutrition Survey (NDNS) database. When such information becomes available, it could be used to provide an indication or estimation of users' exposures to any allergenic proteins in chitin or chitosan-based BBFCMs.

64. Following a full assessment (which is pending additional data and an FSA/FSS risk management decision on a threshold level for crustacea), a statement on chitosan in bio-based food contact materials will subsequently be produced.

## **List of Abbreviations**

ADI      acceptable daily intake

BBFCM   bio-based food contact material

Bw      bodyweight

COS      chitooligosaccharide

COT      Committee on Toxicity of Chemicals in Food, Consumer Products and the Environment

DBPCFC   double-blind, placebo-controlled food challenge

dm<sup>2</sup>      square decimeter

ED      eliciting dose

EFSA      European Food Safety Authority

ELISA      enzyme-linked immunosorbent assay

EU      European Union

FAO      Food and Agriculture Organization of the United Nations

FCM      food contact material

FSA Food Standards Agency

FSANZ Food Standards Australia New Zealand

FSS Food Standards Scotland

GRAS generally recognised as safe

GWP global warming potential

IgE immunoglobulin E

ISO International Organization for Standardization

KDa kilodaltons

KFDA Korean Food and Drug Administration

LCA life-cycle assessment

NAC National Aspergillosis Centre

NDNS national diet and nutrition survey

NF novel food

OML overall migration limit

pl isoelectric point

PEF product environmental footprint



PLA poly(lactic) acid

PM particulate matter

Ppb parts per billion

RfD reference dose

SML specific migration limit

SPT skin prick test

US EPA United States Environmental Protection Agency

US FDA United States Food and Drug Administration

VOC volatile organic compounds

WHO World Health Organization

## References

Barenes H., Phimmasane M., & Rajaonarivo C. (2015) Insect consumption to address undernutrition, a national survey on the prevalence of insect consumption among adults and vendors in Laos. PLoS ONE 10(8): e0136458.

Bradley E.L. (2010). FSA Project A03070: Biobased materials used in food contact applications: an assessment of the migration potential:

[Research report: Biobased materials used in food contact applications: an assessment of the migration potential.](#)

Broekman H.C., Knulst A.C., den Hartog Jager S., et al. (2015) Effect of thermal processing on mealworm allergenicity. Molecular Nutrition and Food Research **59**: 1855-1864.

Broekman H., Verhoeckx K.C., den Hartog Jager C.F., et al. (2016) Majority of shrimp-allergic patients are allergic to mealworm. *J. Allergy Clin. Immunol.* 137: 1261-1263.

Broekman H.C., Knulst A.C., den Hartog Jager C.F., et al. (2017) Primary respiratory and food allergy to mealworm. *Journal of Allergy and Clinical Immunology* **140**: 600-603.

Bueter C.L., Lee C.K., Rathinam V.A., et al. (2011) Chitosan but not chitin activates the inflammasome by a mechanism dependent upon phagocytosis. *J Biol Chem* **286**: 35447-35455.

Boot R.G., Blommaart E.F., Swart E., et al. (2001). Identification of a novel acid mammalian chitinase distinct from chitotriosidase. *Journal of Biological Chemistry* 276: 6770-6778.

Campos C.A., Gerschenson L.N., & Flores S.K. (2011). Development of Edible Films and Coatings with Antimicrobial Activity. *Food Bioprocess Technol.* 4: 849-875.

Chae S.Y., Jang M.K., & Nah J.W. (2005) Influence of molecular weight on oral absorption of water soluble chitosans. *J Control Release.* **102(2)**: 383-394.

Cleenewerck M.B., Martin P., & Laurent D. (1994). Allergic contact dermatitis due to a moisturizing body cream with chitin. *Contact Dermatitis* 31(3): 196-197.

Conrado B., Lerodiakonou D., Gowland M.H., et al. (2021) Food anaphylaxis in the United Kingdom: analysis of national data, 1998-2018. *BMJ* 372: 251.

Daul C.B., Slattery M., Reese G., et al. (1994) Identification of the major brown shrimp (*Penaeus aztecus*) allergen as the muscle protein tropomyosin. *Int. Arch. Allergy Immunol.* 105: 49-55.

Deters A., Petereit F., Schmidgall J., et al., (2008) N-acetyl-d-glucosamineoligosaccharides induce mucin secretion from colonic tissue and inducedifferentiation of human keratinocytes, *J. Pharm. Pharmacol.* **60**: 197-204.

Du W.L., Niu S.S., Xu Y.L., et al. (2009) Antibacterial activity of chitosan tripolyphosphate nanoparticles loaded with various metal ions. *Carbohydr Polym.* **75(3)**: 385-389.

EFSA (2010) Scientific Opinion on the safety of 'Chitin-glucan' as a Novel Food ingredient. *EFSA Journal* **8(7)**: 1687:

[Scientific Opinion on the safety of 'Chitin-glucan' as a Novel Food ingredient - - 2010 - EFSA Journal - Wiley Online Library.](#)

EFSA (2011) Scientific Opinion on the substantiation of health claims related to chitosan and reduction in body weight (ID 679, 1499), maintenance of normal blood LDL-cholesterol concentrations (ID 4663), reduction of intestinal transit time (ID 4664) and reduction of inflammation (ID 1985) pursuant to Article 13(1) of Regulation (EC) No 1924/2006 EFSA Journal **9(6)**: 2214: [Scientific Opinion on the substantiation of health claims related to chitosan and reduction in body weight \(ID 679, 1499\), maintenance of normal blood LDL-cholesterol concentrations \(ID 4663\), reduction of intestinal transit time \(ID 4664\) and reduction of inflammation \(ID 1985\) pursuant to Article 13\(1\) of Regulation \(EC\) No 1924/2006 | EFSA \(europa.eu\).](#)

EFSA (2014) Scientific Opinion on the evaluation of allergenic foods and food ingredients for labelling purposes. EFSA Journal **12(11)**: 3894:

[Scientific Opinion on the evaluation of allergenic foods and food ingredients for labelling purposes | EFSA \(europa.eu\).](#)

EFSA (2020) Safety of dried yellow mealworm (*Tenebrio molitor* larva)

as a novel food pursuant to Regulation (EU) 2015/2283. EFSA Journal 19(1): 6343: [Safety of dried yellow mealworm \(\*Tenebrio molitor\* larva\) as a novel food pursuant to Regulation \(EU\) 2015/2283 | EFSA \(europa.eu\).](#)

Fabec B., Hellstrom T., Henrysdotter G., et al. (2000). Active and intelligent food Packaging. A Nordic report on the legislative aspects. Nordic co-operation, p.29: [Active and Intelligent Food Packaging: A Nordic Report on the Legislative ... - Google Books.](#)

Fajardo P., Martins J.T., Fuciños C., et al. (2010) Evaluation of a chitosan-based edible film as carrier of natamycin to improve the storability of Saloio cheese. J. Food Eng. **101**: 349-356.

FAO/ WHO (2023) Risk assessment of food allergens – Part 2: Review and establish threshold levels in foods for the priority allergens. Meeting Report. Food Safety and Quality Series No. 15. Rome: [Risk Assessment of Food Allergens. Part 2: Review and establish threshold levels in foods for the priority allergens \(fao.org\).](#)

- Ferreira A.R., Alves V.D., & Coelho I.M. (2016) Polysaccharide-based membranes in food packaging applications. *Membranes (Basel)* **6:2**.
- FSANZ (2012) Supporting Document 1: Risk and Technical Assessment Report – Application A1077 Fungal Chitosan as a Processing Aid: [A1077-ChitosanAppR-SD1.pdf \(foodstandards.gov.au\)](#).
- Gades M.D. & Stern J.S. (2003) Chitosan supplementation and fecal fat excretion in men. *Obes Res.* **11(5)**: 683-688.
- Gades M.D. & Stern J.S. (2005) Chitosan supplementation and fat absorption in men and women. *J Am Diet Assoc.* **105(1)**: 72-77.
- Garino C., Mielke H., Knüppel S., et al. (2020) Quantitative allergenicity risk assessment of food products containing yellow mealworm (*Tenebrio molitor*). *Food and Chemical Toxicology* **142**: 111460.
- Han W., Yu Y., Li J., et al. (2011). Application and safety assessment for nano-composite materials in food packaging. *Chinese Science Bulletin* **56(12)**: 1216-1225.
- Hoff M., Trüeb R.M., Ballmer-Weber B.K., et al. (2003) Immediate-type hypersensitivity reaction to ingestion of mycoprotein (Quorn) in a patient allergic to molds caused by acidic ribosomal protein P2. *J Allergy Clin Immunol.* **111(5)**: 1106-10.
- Houben G.F., Baumert J.L., Blom W.M., et al. (2020) Full range of population eliciting dose values for 14 priority allergenic foods and recommendations for use in risk characterization. *Food and Chemical Toxicology* **146**: 111831.
- Huang B., Xiao D., Tan B., et al., (2016) Chitosan oligosaccharide reduces intestinal inflammation that involves calcium-sensing receptor (CaSR) activation in lipopolysaccharide(LPS)-challenged piglets. *J. Agric. Food Chem.* **64**: 245-252.
- Ifuku S. & H. Saimoto (2012) Chitin nanofibers: preparations, modifications, and applications. *Nanoscale* **4(11)**: 3308-18.
- JFCRF (2011) Japan Food Chemical Research Foundation. List of Existing Food Additives: [The Japan Food Chemical Research Foundation \(ffcr.or.jp\)](#).
- Jeevahan J. & Chandrasekaran M. (2019) Nanoedible films for food packaging: a review. *J. Mater. Sci.* **54**: 12290-12318.

- Kato Y., Yagami A., & Matsunaga K. (2005) A case of anaphylaxis caused by the health food chitosan. *Arerugi* **54**: 1427-1429.
- Kim D., Beck B.R., Heo S.B., et al. (2013) *Lactococcus lactis* BFE920 activates the innate immune system of olive flounder (*Paralichthys olivaceus*), resulting in protection against *Streptococcus iniae* infection and enhancing feed efficiency and weight gain in large-scale field studies. *Fish. Shellfish Immunol.* **35**: 1585-1590.
- Kim K.M., Son J.H., Kim S.K., et al. (2006). Properties of chitosan films as a function of pH and solvent type. *Journal of Food Science* 71(3): E119-E124.
- Konovalova M.V., Markov P.A., Durnev E.A., et al. (2017) Preparation and biocompatibility evaluation of pectin and chitosan cryogels for biomedical application. *J Biomed Mater Res.* **105(2)**: 547-556.
- Lodhi G., Kim Y-S., Hwang J-W., et al. (2014) Chitooligosaccharide and Its Derivatives: Preparation and Biological Applications. *BioMed Research International* Article ID 654913.
- Lopata A.L., O'Hehir R.E., & Lehrer S.B. (2010) Shellfish allergy. *Clinical & Experimental Allergy* **40**: 850-858.
- Madhumathi K., Shalumon K.T., Rani V.V., et al. (2009) Wet chemical synthesis of chitosan hydrogel-hydroxyapatite composite membranes for tissue engineering applications. *Int J Biol Macromol.* **45(1)**: 12-15.
- Moraru C., Mincea M.M., Frandes M., et al. (2018) A Meta-Analysis on Randomised Controlled Clinical Trials Evaluating the Effect of the Dietary Supplement Chitosan on Weight Loss, Lipid Parameters and Blood Pressure. *Medicina* 54: 109.
- Nguyen M.X.H. (2012) Characterization of allergenic and antimicrobial properties of chitin and chitosan and formulation of chitosan-based edible film for instant food casing. Melbourne, Australia: Royal Melbourne Institute of Technology (RMIT) University, PhD thesis: [core.ac.uk/download/pdf/15625643.pdf](http://core.ac.uk/download/pdf/15625643.pdf).
- NTP (2017) Technical Report on the Toxicity Study of Chitosan (CASRN 9012-76-4) Administered in Feed to Sprague Dawley [CrI:CD(SD)] Rats. Toxicity Report 93, National Toxicology Program, Public Health Service, U.S. Department of Health and Human Services.
- Ouattara B., Simard R.E., Piette G., et al. (2000) Inhibition of surface spoilage bacteria in processed meats by application of antimicrobial films prepared with

chitosan. *Int. J. Food Microbiol.* **62**: 139-148.

Pereira F., Pereira C., & Lacerda M.H. (1998). Contact dermatitis due to a cream containing chitin and a Carbitol. *Contact Dermatitis* 38(5): 290-291.

Pereira B., Venter C., Grundy J., et al. (2005) Prevalence of sensitization to food allergens, reported adverse reaction to foods, food avoidance, and food hypersensitivity among teenagers. *J. Allergy Clin. Immunol.* 116: 884-892.

Qin C., Li H., Liu Y., et al. (2006) Water-solubility of chitosan and its antimicrobial activity. *Carbohydrate Polymers* **63**: 367-374.

Qin Y., Zhang S., Yu J., et al. (2016) Effects of chitin nano-whiskers on the antibacterial and physicochemical properties of maize starch films. *Carbohydrate Polymers* 147: 372-378.

Reese G., Ayuso R., & Lehrer S.B. (1999) Tropomyosin: An Invertebrate Pan-Allergen. *International Archives of Allergy and Immunology* 119(4): 247-258.

Remington B.C., Westerhout J., Meima M.Y., et al. (2020) Updated population minimal eliciting dose distributions for use in risk assessment of 14 priority food allergens. *Food and Chemical Toxicology* **139**: 111259.

Romano P., Fabritius H., & Raabe D. (2007). The exoskeleton of the lobster *Homarus americanus* as an example of a smart anisotropic biological material. *Acta Biomaterialia* **3(3)**: 301-309.

Sánchez B., González-Tejedo C., Ruas-Madiedo P., et al. (2011) Lactobacillus plantarum extracellular chitin-binding protein and its role in the interaction between chitin, Caco-2 cells, and mucin. *Appl. Environ. Microbiol.* **77(3)**: 1123-6.

Singla A.K. & Chawla M. (2001) Chitosan: some pharmaceutical and biological aspects – an update. *J Pharm Pharmacol* **53**: 1047-1067.

Sinha S., Tripathi P., & Chand S. (2012). A new bifunctional chitosanase enzyme from *Streptomyces* sp. and its application in production of antioxidant chitooligosaccharides. *Applied Biochemistry and Biotechnology* **167**: 1029-1039.

Tapola N.S., Lyyra M.L., Kolehmainen R.M. et al. (2008) Safety aspects and cholesterol-lowering efficacy of chitosan tablets. *J Am Coll Nutr.* **27(1)**: 22-30.

Taylor G. & Wang N. (2018) Entomophagy and allergies: a study of the prevalence of entomophagy and related allergies in a population living in North-Eastern

Thailand. Bioscience Horizons 11(8).

Tee R.D., Gordon D.J., Welch J.A., et al. (1993) Investigation of possible adverse allergic reactions to mycoprotein ('Quorn'). Clin Exp Allergy 23: 257-60.

Tharanathan R.N. (2003). Biodegradable films and composite coatings: Past, present and future. Trends in Food Science and Technology 14(3): 71-78.

TNO (Netherlands Organisation for Applied Scientific Research) (2009). Repeated-dose (13-week) oral toxicity study in rats with chitin-glucan. Study Report provided to EFSA by Kitozyme.

Todorov S.D., Leblanc J.G., Franco B., et al., (2012) Evaluation of the probiotic potential and effect of encapsulation on survival for Lactobacillus plantarum ST16 Pa isolated from papaya. World J. Microbiol Biotechnol **28**: 973-984.

US FDA (2008) HemCon Notification: [K080818.pdf \(fda.gov\)](#).

US FDA (2012) Nutrition Center for Food Safety Applied. GRAS Notice Inventory - Agency Response Letter GRAS Notice No. GRN 000443: [GRAS Notice 000443: Shrimp-derived chitosan \(archive-it.org\)](#) (accessed 01/08/2022).

Van den Broek L.A.M., Knoop R.J.I, Kappen F.H.J., et al. (2015) Chitosan films and blends for packaging material. Carbohydrate Polymers 116: 237-242.

Vasile C. (2018) Polymeric Nanocomposites and Nanocoatings for Food Packaging: A Review. Materials 11: 1834.

Waibel K.H., Haney B., Moore M., et al. (2011) Safety of chitosan bandages in shellfish allergic patients. Military Medicine **176**: 1153-6.

Woo C.K. & Bahna S.L. (2011) Not all shellfish "allergy" is allergy! Clinical and Translational Allergy **1**: 3.

Yadav M., Goswami P., Paritosh K., et al. (2019) Seafood waste: a source for preparation of commercially employable chitin/chitosan materials. Bioresources and Bioprocessing **6**: 8.

Yin M., Lin X., Ren T., et al. (2018) Cytocompatible quaternized carboxymethyl chitosan/poly(vinyl alcohol) blend film loaded copper for antibacterial application. Int J Biol Macromol **120**: 992-998.

Zeng J.B., He Y.S., Li S.L., et al. (2012) Chitin whiskers: an overview. *Biomacromolecules* 13(1): 1-11.